



## Ameliorative Effect of Ascorbic Acid on Hematological Parameters Due to Cadmium Toxicity in *Cyprinus carpio*

Hamza G. Al-Agidi, Kadhim O.M. AL-Humairi and Sadeq M. Al-Haider<sup>1</sup>

Al-Furat AL-Awsat Technical University, AL- Musaib Technical College, 51009, Babylon, Iraq

<sup>1</sup>College of Veterinary Medicine, Al-Qasim Green University, Iraq

E-mail: com.hamza@atu.edu.iq, com.kdm@atu.edu.iq

**Abstract:** The present study showed the protective effects of Vitamin C by reducing the toxicity of cadmium on the hematological parameters in common carp during exposure period. The results of present study showed significant differences at  $P \leq 0.05$  between treatments with cadmium and control, ascorbic acid only and ascorbic acid with cadmium on hematological parameters. The results also showed the effect of Vitamin C (ascorbic acid) to reduce cadmium toxicity after 7 to 14 days after chronic exposure to  $\text{CdCl}_2$ , increased white blood cells number, mean corpuscular volume, Mean Corpuscular Haemoglobin and decreased RBC, Hb content, PCV and the mean corpuscular hemoglobin concentration as compared to control were found. The exposure of cadmium with ascorbic acid the WBC, RBC decreased and Hb contents, RBC, the packed cell volume PCV increase values of these parameters as that in control groups when compared with cadmium exposure groups.

**Keywords:** Ascorbic acid, Cadmium toxicity, Common carp, Hematological parameters

The Heavy metals concentration trend to increase more and more in the all environment spheres especially in the aquatic environment. Fishes have been recognized as good accumulators of organic and inorganic pollutants (Eneji et al 2011). Aquatic organisms such as fish accumulate metals from water or sediment with concentrations higher many times than the concentrations of these metals in the water, also they can concentrate metals at different levels in different organs, and these heavy metals acquired by the chain of food as a result of pollution will be posed chemical hazards and threatening to human. However, low levels of heavy metals, such as copper, cobalt, zinc, iron and manganese are crucial for activity of enzyme activity and many other biological processes. Other heavy metals, for example cadmium and lead are not recognized to have essential role in living organisms and they are toxic even at low concentrations, while the essential metals also developed to become toxic at high level of concentrations, the consequences of heavy metal pollution can be hazardous to humans through food (Sani 2011).

Fish is the most susceptible aquatic animals to the heavy metals. The physical and chemical parameters of the environment in which the fish resides appeared to affect the rate of bioaccumulation of trace elements (Cao et al 2010). Normally, fish may be considered as one of the healthiest and cheapest source of protein due to its higher cysteine amino acid composition than most other sources of protein. The effects of exposure to any hazardous constituent depend on the dose, exposure time, exposure mode, and presence of

other chemicals (Adekola et al 2007). Therefore, heavy metals monitoring in aquatic environment are very important, especially for breeding of fish. Increased loads of toxic metals in waste water may increase the risk of water contamination in the current study, the hematological parameters were studied to evaluate the cadmium chloride impacts on carp fish.

Red blood corpuscle RBC and hemoglobin can be used as an indicator for anemia, fluid volume disturbances. In general, heavy metals have acute effect in hematological parameters and cause reduction in RBC and Hb. Further, anemia could be happened due to plasma increasing volume caused by disturbed water balance and will decrease RBC production (Saraswathi et al 2003). In addition, as the haemoglobin (Hb) is the main constituents of blood, which transport of oxygen throughout the blood capillaries, decreasing the haemoglobin level might reduce the biological activity of fishes due to stress syndrome. Researchers have reported alteration in RBC and Hb level (Garg et al 1989, Magare and Patil 2000), and also in the present investigation on the protective effect of ascorbic acid to reduce the toxicity of cadmium on hematological parameters in *Cyprinus carpio*.

### MATERIAL AND METHODS

**Experimental fishes:** The freshwater common carp *Cyprinus carpio* with a size range of 16-20cm and, weighing  $54 \pm 4$  g irrespective of their sex, have been chosen as the test organism in the present study. Before the exposure

experiment fishes were acclimatized to the laboratory conditions in large fiber glass tanks with unchlorinated ground water for 3 days at a room temperature of  $28 \pm 20^\circ\text{C}$ . In order to avoiding the carp fishes overcrowding as its benthic in nature, five fishes putted in each tank. After every 24h, water was changed completely. To prevent the escaping of fishes, tanks were covered with net. No feed used during the experiment period (14 days).

**Chemicals and reagents:** Cadmium chloride ( $\text{CdCl}_2$ ) and vitamin C (L-Ascorbic acid) were obtained from the environmental laboratory at the Collage of Science, Babylon University, then the  $\text{CdCl}_2$  was dissolved in  $\text{H}_2\text{O}$  ( $1 \text{ mg l}^{-1}$ ) and the L-Ascorbic acid was added with a ratio of  $50 \text{ mg l}^{-1}$  (Al Maamouri 2011). The other chemicals (reagent grade) were obtained from local scientific distributors in Iraq.

**Animals and experimental design:** 60 healthy adult individuals of common carp *Cyprinu scarpio* ( $60 \pm 5 \text{ g}$ ) were obtained from a registered fish farm in Babylon province. 3 days period of time was used for fish acclimation. Then it spread in glass aquaria with (dimensions,  $90 \times 30 \times 50 \text{ cm}$ ) filled with 120 L fresh water (de-chlorinated); the carp fishes were kept in a laboratory, setting with a photoperiod, light (12 hours) and dark cycle (12 hours). Furthermore, temperature was maintained of  $28 \pm 2^\circ\text{C}$  and then water aquarium was aerated. Water dissolved oxygen level ( $6.5 \text{ mg l}^{-1}$ ), pH ( $6.9 \pm 0.4$ ), and electrical conductivity ( $219 \pm 2 \mu\text{m ho cm}^{-1}$ ) of aquaria water which measured using multi meter; conditions were closely monitored and kept stable during the experiments. They were subdivided into four groups with 15 individuals for each one. Control group were given tap water only. Second group were given Vitamin C at a dose  $50 \text{ mg l}^{-1}$ . Third groups were given cadmium at a dose  $1 \text{ mg l}^{-1}$  and fourth groups were (Vitamin C and cadmium) given Vitamin C  $50 \text{ mg l}^{-1}$  prior three hours of cadmium in concentration  $1 \text{ mg l}^{-1}$  administration. *Cyprinu scarpio* was exposed to sub-lethal concentration of Cadmium for 14 days. For sub-lethal exposure experiment, three replicates were maintained. The water was changed every day, so that the concentration of cadmium chloride remained the same during the experimental period.

**Haematological studies:** Blood was taken at the end of 7 day and 14 day by catching each fish very gently using a small net, one by one with minimal disturbances. Each fish wrapped in a clean and dry towel was trapped, the posterior half of its body was cleaned with coarse clean paper. The blood samples were collected using severance of caudal peduncle and placed in Eppendorf tubes holding 1% (EDTA) (Mgbenka et al 2003). Haematological parameters were evaluated using the methods of Hesser (1960) and Blaxhall and Daisley (1973).

**Red Blood Corpuscular (RBC) measurement:** Counting of RBC was done with a Neubauer chamber, Sohn and Henry (1969). Red glass bead pipette was used for indicating the counting chamber (triplicate).

**Haemoglobin (Hb) content:** Cyanmethemoglobin method was performed for determination of Hb (Dacie and Lewis 1968). 0.02 ml of blood sample was pipetted into Darbkinp's reagent (5ml) and shaken then allowed to settle for 10 min. after that, absorbance was measured at 540 nm wavelength with spectrophotometer against a reagent blank.

**Estimation of Packed Cell Volume (PCV):** Evaluation of PVC was achieved using micro-haematocrit method (Snieszko 1960). Non-clotted blood specimens (Heparinized) were collected in even bored capillaries. It was indorsed to run 1/2 - 3/4 lengths of capillary tube closed with sealing wax on opposite sides and then assigned to a high speed micro-haematocrit centrifuge (12000 rpm) for 5 min.

**White Blood Corpuscles (WBC) measurement:** WBCs were measure during the method described by Donald Hunter and Bomford (1963).

**Statistical analysis:** Data analysis was done using SPSS ver. 23, and present as mean  $\pm$  SD. It was considered as significant when P value was 0.05.

## RESULTS AND DISCUSSION

The results of presents study were to determine the impact of cadmium toxicity on *C. carpio* and ameliorative effect by L-ascorbic acid on the levels of hematological indices, the hematological indices in presents study showed significant differences at  $p \leq 0.05$  among the treatments. Ascorbic acid alone has no effect on all hematological markers as compared with treatments of cadmium

**RBCs:** Red blood cells number in control was  $3.5$  and  $3.3 \times 10^6 \text{ mm}^{-3}$  at 7 and 14 days respectively. While in cadmium exposure, its number was significantly decreased ( $2.62$ – $1.46$ )  $10^6 \text{ mm}^{-3}$  at 7 and 14 days, respectively as compared with treatment groups with ascorbic acid which its numbers significantly increased at 7 and 14 days ( $3.12$ – $3.3$ )  $10^6 \text{ mm}^{-3}$ , respectively (Fig. 1).

**WBC:** The number of white blood cells in cadmium exposure significantly increased at 7 and 14 days and reached to  $4.32$ – $5.87 \times 10^6 \text{ mm}^{-3}$ . While it was decreased significantly in treatment with ascorbic acid along with cadmium exposure to be ( $3.87$  –  $3.11 \times 10^6 \text{ mm}^{-3}$ ). The WBC for the control treatment was ( $3.22$ – $3.17 \times 10^6 \text{ mm}^{-3}$ ) (Fig. 2).

**Hb:** The Hb level in treatment with ascorbic acid with cadmium exposure reached at 7 and 14 days ( $9.1$ – $8.9 \text{ g } 100 \text{ ml}^{-1}$ ) as compared with control groups at 7 and 14 days ( $9.6$ – $10.8$ )  $\text{g } 100 \text{ ml}^{-1}$ , respectively. The level of hemoglobin percent in cadmium exposure was significantly decreased at

7 and 14 days to reach (8.3-7.4) g 100 ml<sup>-1</sup>(Fig. 3).

**PCV:** The packed cell volume PCV% in the control was (34.6-33.6%) at 7 and 14 days and its percentage in cadmium treatment were significantly decreased from 29.4%-26% and vitamin C+ cadmium treatments were from 32.9-33.8% at 7 and 14 days, respectively (Fig. 4).

The percent of MCV in cadmium exposure was significantly increased at 7 and 14 days and reached to 112.21-178.08 μm<sup>3</sup> While its concentration in treatment with ascorbic acid plus cadmium exposure was significantly decreased at 7 and 14 days (108.33-97.57 μm<sup>3</sup>) as compared with control groups at 7 and 14 days (98.85-108.33 μm<sup>3</sup>), respectively (Fig. 5).

The MCH in cadmium exposure was 31.67-44.8 pg, while in treatment with vitamin C+ cadmium MCH was reached 29.16-31.98 pg and in control was ranged between 30.85-30.35 pg at 7 and 14 days exposure respectively (Fig. 6). The MCHC in cadmium exposure was significantly

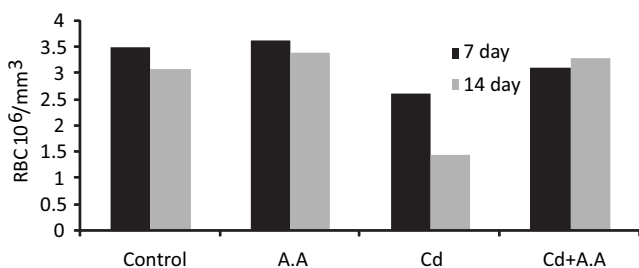


Fig. 1. Red blood corpuscular counts 10<sup>6</sup> mm<sup>-3</sup> in bloods of *C. carpio* during period of exposure

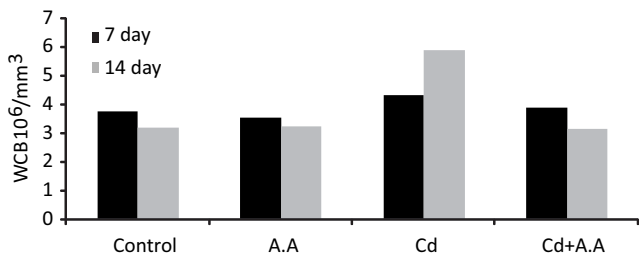


Fig. 2. White blood cells counts 10<sup>6</sup> mm<sup>-3</sup> in bloods of *C. carpio* during exposure period

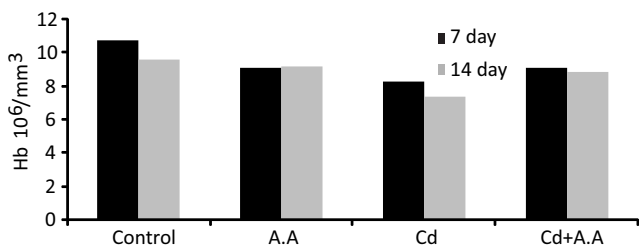


Fig. 3. Hb 10<sup>6</sup> mm<sup>-3</sup> in bloods of *C. carpio* during period of exposure

increased at 7 and 14 days and reached 28.23 and 16.85%, respectively. While its concentration in treatment with ascorbic acid plus cadmium exposure decreased at 7 and 14 days (26.92-26.23%) as compared with control groups at 7 and 14 days (31.21-25.8%), respectively (Fig. 7).

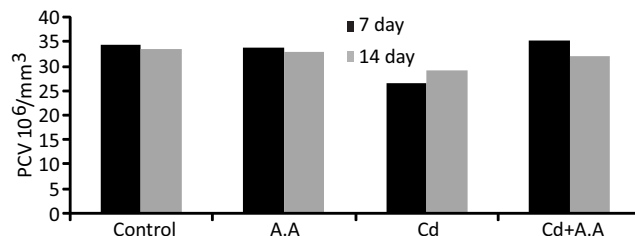


Fig. 4. PCV 10<sup>6</sup> mm<sup>-3</sup> in bloods of *C. carpio* during period of exposure

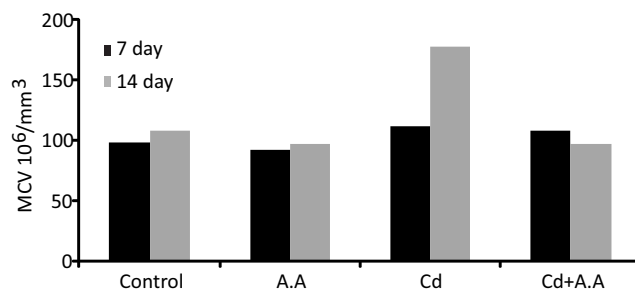


Fig. 5. MCV 10<sup>6</sup> mm<sup>-3</sup> in bloods of *C. carpio* during period of exposure

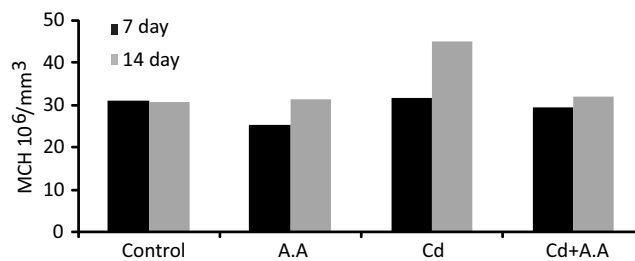


Fig. 6. MCH 10<sup>6</sup> mm<sup>-3</sup> in bloods of *C. carpio* during period of exposure

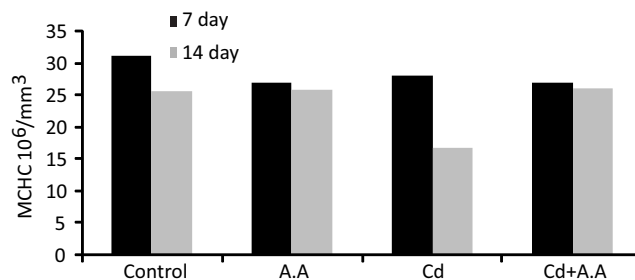


Fig. 7. MCHC% 10<sup>6</sup> mm<sup>-3</sup> in bloods of *C. carpio* during period of exposure

## REFERENCES

- Abbas HHH 2006. Acute toxicity of ammonia to common carp fingerlings (*Cyprinus carpio*) at different pH levels. *Pakistan Journal of Biological Sciences* **9**: 2215-2221.
- Adekola FA and Eletta OAA 2007. A study of heavy metal pollution of Asa River, Ilorin, Nigeria; trace metal monitoring and geochemistry. *Environmental Monitoring and Assessment* **125**(1-3): 157-163.
- Al Maamouri and Anas M Mohamad 2011. *Comparative study of the effects of zinc and chromium elements on common carp fish*. M.Sc. Thesis, Collage of Science- University of Babel.
- Agarwal RA 1983. Pesticides and environmental pollution. *Indian Farming* **33**(5): 21-25.
- Blaxhall PC and Daisley KW 1973. Routine haematological methods for use with fish blood. *Journal of Fish Biology* **5**: 771-781.
- Blaxhall PC 1972. The hematological assessment of the health of freshwater fishes: A review of selected literatures. *Journal of Fish Biology* **4**: 593-604.
- Chauhan RRS, Saxena RK and Kumar S 1994. Rogor induced haematological alterations in *Cyprinu scarpio*. *Adv. Bios.* **13**: 57.
- Christensen GM, Mickim JM, Brings WA and Hunt EP 1972. Changes in the blood of brown bull head, (*Ictalurus nebulosis*) following short and long term exposure to copper (II). *Toxicology and Applied Pharmacology* **23**: 417-427.
- Donald Hunter and Bomford 1963. A dangerous misprint. *British Medical Journal* **1**: 1024.
- Enejil S, Sha'Ato R and Annune PA 2011. Bioaccumulation of heavy metals in fish (*Tillapia zilli* and *Clarias gariepinus*) organs from river Benue, North Central. *Pakistan Journal of Analytical Envnt.* **12**(1&2): 25-31.
- Garg VK, Garg SK and Tyagi SK 1989. Manganese induced haematological and biochemical abnormalities in *Heteropneustes fossilis*. *Journal of Environmental Biology* **10**: 349-353.
- Gill TS, Pant SC and Pant J 1988. Gill, liver and kidney lesions associated with experimental exposure to carbaryl and diemethoate in the fish, *Puntius conchoniuis*. *Bulletin of Environmental Contamination and Toxicology* **41**: 71-78.
- Gupta A, Rai DK, Pandey RS and Sharma B 2009. Analysis of some heavy metals in the riverine water, sediments and fish from river Ganges at Allahabad. *Environment Monitoring Assessment* **157**: 449-458.
- Hesser EF 1960. Methods for routine fish hematology. *The Progressive Fish Culturist* **22**(4): 164-171.
- Kori-Siakpere O, Ake JEG and Idoge E 2005. Haematological characteristic as of the African snakehead, *Parachanna obscura*. *African Journal of Biotechnology* **4**: 527-530.
- Kumar R and Agarwal VP 1993. Effect of sublethal value of LC50 of mercury chloride on haematological parameters on *Clarius batrachus*. *Environment and Ecology* **2**: 23-25.
- Magare SR and Patil HT 2000. Effect of pesticides on oxygen consumption, red blood cell count and metabolites of fish, *Puntius ticto*. *Environment and Ecology* **18**(4): 891-894.
- Mc Donald MD and Grosell M 2006. Maintaining osmotic balance with an aglomerular kidney. *Comparative Biochemistry and Physiology* **143**: 447-458.
- Mgbenka BO, Oluah NS and Umeike I 2003. Effect of Gammalin 20 (lindane) on differential white blood cell counts of the African catfish, *Clarias albopunctatus*. *Bulletin of Environmental Contamination and Toxicology* **71**(2): 248-254.
- Sani U 2011. Determination of some heavy metals concentration in the tissues of Tilapia and Catfishes. *Biokemistri* **23**(2): 73-80.
- Sarswathi K, Ramesh M, Jayalakshmi K, Revathy B and Noortheen A 2003. Haematological responses of fingerlings of *Cyprinus carpio* to sublethal toxicity of sodium nitrite exposure. *Aquaculture* **4**(1): 49-56.
- Sen G, Bahera MK, Behera R and Patel PN 1993. Toxic effect of zinc on freshwater fish, *Channa punctatus*. *Environment and Ecology* **11**: 735-737.
- Shalaby AM, Abbas HHH and Khatab YA 2005. The effect of sublethal doses of cadmium on the disposition of some trace elements and liver function in common carp *Cyprinu scarpio* L. *Egyptian Journal of Basic and Applied Physiology* **4**: 383-395.
- Snieszko SF 1960. Microliacmatocrit as a tool in fisheries management. Special Scientific Reports-Fisheries. No. 314. U. S. Dept. Inter. Fish and Fisheries Wildlife Special Sci-Report, 15.
- Sohn ID and Henry IB 1969. *In: Todd-Sunford clinica diagnosis by Laboratory methods*, 14th edn., W. B. Saunders company, Philadelphia, London, Toronto. **19**: 139-143.
- Zikić RV, Stajin S, Pavlović SZ, Ognjanović BI and Saičić ZS 2001. Activities of superoxide dismutase and catalase in erythrocyte and plasma transaminases of goldfish (*Carassius auratus gibelio Bloch*) exposed to cadmium. *Physiological Research* **50**: 105-111.
- Web JL 1966. Enzymes and metabolic inhibitor. Vol. 3. Academic press. New York.